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Evolutionary analysis of genes associated with the sense of balance in semi-aquatic mammals

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Abstract

Background Semi-aquatic mammals represent a transitional phase in the evolutionary spectrum between terrestrial and aquatic mammals. The sense of balance is crucial for mammalian locomotion, and in semi-aquatic mammals, the structural foundation of this sense (the vestibular system) shows distinct morphological adaptations to both aquatic and terrestrial environments compared to their terrestrial counterparts. Despite this, the precise molecular mechanisms driving these adaptations remain elusive. Our study endeavors to unravel the genetic components associated with the sense of balance in semi-aquatic mammals and to examine the evolutionary trajectories of these genes, shed light on the molecular mechanisms underlying the adaptive evolution of balance perception in semi-aquatic mammals.

Results We selected 42 mammal species across 20 orders, 38 families, and 42 genera for analysis. We analyzed a comprehensive set of 116 genes related to the vestibular system's development or function. Our findings indicate that 27 of these genes likely experienced adaptive evolution in semi-aquatic mammals. Particularly, genes such as *SLC26A2*, *SOX10*, *MYCN*, and *OTX1* are implicated in collectively orchestrating morphological adaptations in the semicircular canals to suit semi-aquatic environments. Additionally, genes associated with otolith development, including *SLC26A2*, *OC90*, and *OTOP1*, likely regulate otolith sensitivity across various locomotor modes. Moreover, genes linked to vestibular disorders, such as *GJB2*, *GJB6*, and *USH1C*, may provide a molecular foundation for averting vertigo amidst intricate locomotor scenarios in semi-aquatic mammals.

Conclusions Our research offers insights into the molecular mechanisms underlying the evolution of the sense of balance in semi-aquatic mammals, while also providing a new research direction for the adaptive evolution of mammals undergoing a secondary transition to an aquatic lifestyle.

Keywords Semi-aquatic mammals, Vestibular system, Sense of balance, Adaptive evolution, Molecular basis

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Background

The sense of balance is vital to the survival of animals, allowing for spatial orientation and stable posture during movement [1]. It is essential for perceiving self-motion, maintaining postural control, and stabilizing during locomotion, often considered the animal's sixth sense [2, 3]. The vestibular system serves as the anatomical cornerstone for generating, perceiving, and exercising balance. A part of this system comprises three semicircular



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canals (the anterior, posterior, and horizontal canals) that detect rotational movements in all directions Another component of the vestibular system includes two otolith organs—the utricle and the saccule, which are positioned horizontally and vertically, respectively, and are responsible for detecting linear acceleration and head tilt in the horizontal and vertical planes [1, 2, 4].

The vestibular system is crucial for maintaining balance and has undergone significant evolutionary selection related to locomotion [5]. Specifically, the size and morphological variability of the semicircular canals within the bony labyrinth, such as roundness, orthogonality, and aspect ratio, are closely associated with locomotor mode [1, 6]. A comparative study of these canals in mammals with diverse locomotor behaviors, including burrowing, arboreal, slithering, and subterranean living, reveals a correlation between canal morphology and locomotor modes. Moreover, there is a significant difference in the declination angle of the semicircular canals between subterranean-living taxa and slithering taxa [7].

With the secondary adaptation of certain vertebrates to aquatic environments, their vestibular system undergoes alterations, leading to distinctions from their terrestrial relatives. A morphological comparison of semicircular canals in musteloids across different habitats showed notable differences between semi-aquatic and terrestrial species. Semi-aquatic otters and minks exhibit elliptical anterior semicircular canals, increased curvature of the horizontal semicircular canals, and reduced angles between posterior and horizontal canals. All of these morphological features correlate with the animal's sensitivity to detecting head movements [8]. Cetaceans, have uniquely small semicircular canal arcs relative to body weight, thus reducing the sensitivity of their vestibular system and likely matching their rapid body-rotation behavior [9]. Similarly, aquatic manatees and semiaquatic seals exhibit reduced semicircular canals [10-12]. Since the Mesozoic era, numerous reptiles transitioned from terrestrial to aquatic habitats, including the extinct thalattosuchians. Marine-dwelling thalattosuchians species displayed smaller labyrinth, wider semicircular canal diameters, and enlarged vestibules akin to those seen in other marine reptiles and cetaceans, aiding in swimming [11]. The canal shape variations in amniotes also suggest a pattern linked to the transition from terrestrial to aquatic locomotion. Physical differences exist between air and water, so the vestibular system differs in terrestrial and aquatic species [13].

The vestibular system is essential for balance and spatial orientation, and extensive research using mice and zebrafish has investigated its genetic basis. These studies seek to understand how genes influence vestibular development and behavior, as well as the molecular causes of vestibular disorders in humans [4, 14, 15].Proper development of the vestibular system relies on the precise temporal expression and regulation of various genes [16]. Gene deletions or mutations can disrupt semicircular canal formation, as seen in mice with BMP2 gene knockouts lacking all three canals [17], or the absence of specific canals due to mutations in genes like *OTX1* [18] and *FGF10* [19]. Furthermore, specific genes play a role in the formation of otoliths, hair cells, and other vestibular structures. For example, mutations in *SLC26A4* [20] and *ATP2B2* (*PMCA2*) [21] affect otolith structure. Similarly, mutations in *CELSR1* [22] and *MITF* [23] impact vestibular hair cell growth, with the latter specifically influencing saccule hair cells. The comprehensive role of genes in vestibular development is highlighted by the abnor-

Although the structural features of the vestibular system in semi-aquatic mammals differ significantly from those of terrestrial animals, the genes that are dependent on the normal development of the vestibular system have been studied extensively. Nonetheless, the molecular underpinnings behind these structural disparities remain enigmatic. Therefore, we curated a gene set linked to the sense of balance and conducted evolutionary analyses to elucidate the molecular basis of the modified vestibular system features in semi-aquatic mammals.

malities in otoliths, semicircular canals, and hair cells in

zebrafish embryos lacking the HDAC1 gene [24].

Results

Sequences retrieval of genes associated with the sense of balance

We selected 12 semi-aquatic mammals and 30 terrestrial mammals across the entire mammalian range as our research subjects. These 42 species belong to 42 genera, 38 families, and 20 orders (Table 1). Through literature search, we compiled a set of 116 genes related to the sense of balance (refer to Additional file 1: Table S1 for the gene list and corresponding references). Based on the constructed gene sets, an extensive collection of 4731 sequences was obtained from 42 mammalian species. Among these, 3746 sequences were directly retrieved from the NCBI database (refer to Additional file 1: Table S2 for the respective sequence search version numbers). Furthermore, a total of 985 sequences were retrieved through a comprehensive BLAST search, encompassing unannotated genomes as well as target sequences that were not previously explored within annotated genomes (see Additional file 2 for detailed raw data of all sequences). Finally, through cluster analysis and bidirectional BLAST analysis based on combinations of datasets from different species, we confirmed that all obtained sequences are orthologous genes of the 116 genes (Additional file 3).

Table 1 List of 42 mammals selected for this study

Order	Family	Genus	Species	Abbreviation	
Monotremata	Tachyglossidae	Tachyglossus	Tachyglossus aculeatus	Tacu	
Didelphimorphia	Didelphidae	Monodelphis Monodelphis domestica		Mdom	
Dasyuromorphia	Dasyuridae	Sarcophilus	Sarcophilus Sarcophilus harrisii		
Diprotodontia	Phascolarctidae	Phascolarctos	Phascolarctos cinereus	Pcin	
	Phalangeridae	Trichosurus	Trichosurus vulpecula	Tvul	
Cingulata	Dasypodidae	Dasypus	Dasypus novemcinctus	Dnov	
Pilosa	Megalonychidae	Choloepus	Choloepus didactylus	Cdid	
Hyracoidea	Procaviidae	Procavia	Procavia capensis	Рсар	
Proboscidea	Elephantidae	Elephas	Elephas maximus	Emax	
Macroscelidea	Macroscelididae	Elephantulus	Elephantulus edwardii	Eedw	
Tubulidentata	Orycteropodidae	Orycteropus	Orycteropus afer	Oafe	
Afrosoricida	Tenrecidae	Echinops	Echinops telfairi	Etel	
Eulipotyphla	Talpidae	Condylura	Condylura cristata	Ccri	
	Erinaceidae	Erinaceus	Erinaceus europaeus	Eeur	
	Soricidae	Sorex	Sorex araneus	Sara	
Scandentia	Tupaiidae	Tupaia	Tupaia belangeri chinensis	Tbel	
Primates	Hominidae	Ното	Homo sapiens	Hsap	
	Hylobatidae	Nomascus	Nomascus leucogenys	Nleu	
	Cercopithecidae	Macaca	Macaca mulatta	Mmul	
Carnivora	Mustelidae	Enhydra	Enhydra lutris	Elut	
	Mustelidae	Lutra	Lutra lutra	Llut	
	Mustelidae	Aonyx	Aonyx cinereus	Acin	
	Mustelidae	Neovison	Neovison vison	Nvis	
	Mustelidae	Martes	Martes flavigula	Mfla	
	Otariidae	Zalophus	Zalophus californianus	Zcal	
	Odobenidae	Odobenus	Odobenus rosmarus divergens	Oros	
	Phocidae	Phoca	Phoca vitulina	Pvit	
	Ursidae	Ursus	Ursus maritimus	Umar	
	Felidae	Acinonyx	Acinonyx jubatus	Ajub	
	Canidae	Vulpes	Vulpes ferrilata	Vfer	
Perissodactyla	Equidae	Equus	Equus caballus	Ecab	
Pholidota	Manidae	Manis	Manis javanica	Mjav	
Artiodactyla	Hippopotamidae	Hippopotamus	Hippopotamus amphibius	Hamp	
	Suidae	Sus	Sus scrofa	Sscr	
	Bovidae	Bos	Bos taurus	Btau	
Lagomorpha	Leporidae	Oryctolagus	Oryctolagus cuniculus	Ocun	
	Ochotonidae	Ochotona	Ochotona princeps	Opri	
Rodentia	Muridae	Mus	Mus musculus	Mmus	
	Bathyergidae	Heterocephalus	Heterocephalus glaber	Hgla	
	Cricetidae	Mesocricetus	Mesocricetus auratus	Maur	
	Castoridae	Castor	Castor canadensis	Ccan	
	Hydrochaeridae	Hydrochoerus	Hydrochoerus hydrochaeris	Hhyd	

Note: Species scientific names and abbreviations labeled in red indicate that the species is a semi-aquatic mammal; all other species are terrestrial mammals

We used the obtained orthologous genes to construct a phylogenetic tree based on the Maximum Likelihood (ML) method and the Bayesian Inference (BI) method. The results indicate that the topological structures of the phylogenetic trees produced by both methods are consistent with the topological structure obtained from the TimeTree database (http://www.timetree.org/). We have refined the ML tree and presented it (Fig. 1). The subsequent analyses are all based on the topology of this tree.



Fig. 1 Phylogenetic tree of 42 species constructed using the maximum likelihood (ML) method. The ends of the evolutionary branches denote the abbreviated Latin names of the 42 species. The branches highlighted in yellow represent the lineages of semi-aquatic species, with the corresponding animal illustrations indicating the semi-aquatic species investigated in this study, while the other branches represent terrestrial species. The varying background colors of the evolutionary branches indicate the different families to which the species belong. The outermost arcs of different colors represent the different orders to which the species belong. The full names corresponding to the abbreviated Latin names of the species, as well as the details of the families and orders to which they belong, can be found in Table 1

Selection pressure analysis

Using the branch-site model, we successfully identified two genes, *EYA1* and *SLC26A2*, under positive selection in semi-aquatic mammals (Table 2). In heterozygous mice, reducing the expression of the *EYA1* gene to only 21% of the normal level resulted in the absence of vestibular sensory formation. Notably, the *EYA1* gene showed exclusive expression during the differentiation of hair

cells, highlighting its pivotal role in the development of vestibular sensory and hair cells [25]. Similarly, when the *SLC26A2* gene was knocked down in zebrafish, notable abnormalities in the morphology of otolith and semicircular canals occurred due to apoptosis, leading to imbalanced swimming behavior [26].

Recognizing the potential for positive selection in genes related to balance in terrestrial mammals, we

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Table 2	List of	nositivel	/ selected	denes in	semi-ad	illatic arolli	n mammals	(branch-site model)	
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Gene Name	Model	InL	2∆lnL	p_Value	Level Parameters	Positively selected sites (BEB)
EYA1	ma	-12,352.896			w0=0.029, w1=1.000, w2=20.619	175 S 0.981*
	ma0	-12,359.559	13.327	< 0.001	w0=0.029, w1=1.000, w2=1.000	
SLC26A2	ma	-21,685.825			w0=0.066, w1=1.000, w2=3.630	22 0.901; 492 E 0.885; 582 A 0.932; 593 T 0.865
	ma0	-21,687.810	3.970	0.046	w0=0.066, w1=1.000, w2=1.000	

Note: InL represents log-likelihood values, and the * symbol represents posterior probability (BEB) greater than 95%

performed an extensive site model analysis on all terrestrial species using two distinct methods (comparing between the M8 and M7 models, and the M2 and M1 models). The results of the two methods (Additional file 1: Table S3 and S4) showed that a total of 16 genes exhibited significant positive selection signals in terrestrial group species but did not contain positively selected genes in the semi-aquatic group, indicating that the two genes detected in semi-aquatic mammals were specifically favored for their adaptation to the semi-aquatic mode of locomotion.

Divergent evolution genes are characterized by their experience of significantly distinct selection pressures across various evolutionary clades. To compare the evolutionary patterns of genes related to sense of balance in semi-aquatic and terrestrial groups, we utilized the Clade model C (CmC) for our analysis. The results demonstrated that 17 genes in the semi-aquatic group were classified as divergent evolution genes (Additional file 1: Table S5). In addition to the SLC26A2 gene identified through positive selection, several other genes, including CASP3 [27], COG4 [28], OTX1 [18], SOX10 [29], and TSKU [30], are involved in the formation of the semicircular canals. Genes like NOXO1 [31], OC90 [32], OTOP1 [33], and SLC26A4 [20] contribute to the formation of otoliths. The absence of the NEUROD1 gene leads to abnormalities in vestibular function [34]. The divergent evolution gene WNT3A, along with related genes like WNT1, plays a crucial role in promoting the development of the vestibule in the inner ear. Interestingly, double mutant mouse embryos lacking both WNT3A and WNT1 genes exhibit a complete absence of vestibular structures [35]. Abnormalities in divergent evolution gene HES1 and related genes like HES5 result in an overproduction of hair cells in the utricle and saccule [36]. Furthermore, the CDH23 [37], GJB6 [38], KCNA10 [39], and USH1C [40] genes have been linked to various types of human vestibular dysfunction disorders.

Rapidly evolved genes are protein-coding genes that undergo a higher rate of amino acid substitutions between species than average. These genes, along with positively selected genes, often play a key role in adaptive evolution in response to changing environments [41, 42]. In this study, we identified 11 genes that are undergoing rapid evolution according to the Branch model (Additional file 1: Table S6). Of these, eight were also detected by the branch-site model and Clade model C. Furthermore, the rapidly evolved genes *ADGRG6* (*GPR126*) [43], *HDAC1* [24], and *ILDR1* [44] are implicated in significant roles during the development of the semicircular canals.

Identification of genes with correlations between evolutionary rates and habitats

In phylogenetics, where each gene may demonstrate variable evolutionary rates across species, establishing correlations between gene evolutionary rates and diverse movement patterns necessitates thorough consideration. To mitigate the influence of phylogenetic factors on gene evolutionary rates, we employed the Phylogenetic Independent Contrast (PIC) approach. Our investigations revealed significant correlations between evolutionary rates and locomotor modes for 10 genes (Fig. 2). Among these, SOX10, GATA2 [45], and MYCN [46] genes are associated with semicircular canal formation, while OC90, OTOP1, and SIX1 [47] genes are linked to otolith formation. Furthermore, the SLC26A2 gene is implicated in both semicircular canal and otolith formation, whereas GJB2 [38], GJB6, and USH1C genes are associated with vestibular disorder.

Identification of convergent evolution genes

Different clades of semi-aquatic mammals may have undergone similar molecular adaptations to facilitate their transition to aquatic locomotion. We considered the likelihood of multiple amino acids with comparable biochemical properties conferring similar fitness at a given locus, we employed the PCOC (Profile Change with One Change) method to identify signals of convergent evolution by detecting shifts in amino acid properties. Our analysis revealed convergent evolution in six genes, each exhibiting changes in amino acid properties at least one locus in semi-aquatic species (Fig. 3). These genes include, in addition to those previously mentioned (OTOP1 and SLC26A2), CHD7, which plays a crucial role in the development of semicircular canals. The complete deletion of CHD7 results in hypoplasia of the semicircular canals and vestibular sensory organs [48]. Furthermore, the GSDME (DFNA5) gene, which also influences semicircular canal development [49], showed convergent evolution. Specific convergent sites of amino acid changes include amino acid 2152 of CHD7, amino acids 236 and 650 of GSDME, amino acid 263 of OTOP1, and amino acids 404 and 744 of SLC26A2.

Overview of different gene sets

To investigate the evolutionary patterns of genes associated with the sense of balance in mammals transitioning to a semi-aquatic locomotor mode, we delved into 27 genes implicated in maintaining balance during semi-aquatic adaptation. This gene set consisted of two positively selected genes, 17 divergent evolution genes, 11 rapidly evolved genes, 10 genes correlating



Fig. 2 Genes with significant correlation between evolutionary rate and habitat type. (**A**)—(**J**) represent the genes *GATA2*, *GJB2*, *GJB6*, *MYCN*, *OC90*, *OTOP1*, *SIX1*, *SLC26A2*, *SOX10*, and *USH1C*, respectively. The horizontal axis represents habitat types corresponding to different modes of locomotion, that is, the contrast between semi-aquatic and terrestrial habitats. The vertical axis represents the contrast of evolutionary rates of genes in species with different modes of locomotion, with values transformed by log10. All *p* values for the correlations are less than 0.05



Fig. 3 Genes with convergent substitutions of amino acids exhibiting similar biochemical characteristics in semi-aquatic mammals. The left side of the figure shows a phylogenetic tree of 42 species, with the ends of the branches representing their abbreviated Latin names, corresponding to the full names listed in Table 1. Yellow highlights denote branches of semi-aquatic mammals, while black vertical lines demarcate the semi-aquatic species into different evolutionary clades, indicating independently occurring secondary aquatic adaptations. The right side lists the amino acid positions with convergent substitutions: the 2152nd of *CHD7*, the 236th and 650th of *GSDME*, the 263rd of *OTOP1*, and the 404th and 744th of *SLC26A2*. An "×" with no background color indicates that the gene was not identified in this species, and this evolutionary branch was excluded from the analysis of that gene. The posterior probabilities for each convergent site are all greater than 0.95

with semi-aquatic habitat, and six genes exhibiting convergent evolution (Fig. 4). Notably, the *SLC26A2* gene, crucial for the normal development of otoliths and semicircular canals, was consistently identified across all five datasets. Additionally, *OTOP1* was present in four datasets — divergent evolution genes, rapidly evolved genes, genes correlating with semi-aquatic habitat, and genes undergoing convergent evolution. Furthermore, the *GJB6* and *USH1C* genes were identified by all three methods, whereas the *CASP3*, *CDH23*, *KCNA10*, *OTX1*, *OC90*, and *SOX10* genes were concurrently detected by two methods. These ten genes are proposed as promising



Fig. 4 Venn diagram showing the overlap between positively selected genes, divergent evolution genes, rapidly evolved genes, genes with semi-aquatic correlation, and convergent evolution genes. The *SLC26A2* gene is included in all five datasets. The *OTOP1* gene is present in the genes with correlation, convergent evolution genes, divergent evolution genes, and rapidly evolved genes. The *GJB6*, *USH1C*, *OC90*, *SOX10*, *CASP3*, *CDH23*, *KCNA10* and *OTX1* genes have also been detected by at least two methods

candidates that underwent adaptive evolution in semiaquatic mammals to maintain locomotor balance during terrestrial ambulation and aquatic swimming.

To elucidate interactions among the 27 genes from the five datasets, we cross-referenced them with the STRING database. The analysis results indicate that there are significant interactions among the proteins of 21 genes (p < 1.0e-16) (Fig. 5), and they are significantly enriched in GO terms such as semicircular canal development (GO:0060872, FDR=0.0002) and semicircular canal morphogenesis (GO:0048752, FDR=0.0092).

Discussion

To comprehensively investigate the evolutionary patterns of genes associated with the sense of balance in semi-aquatic mammals and uncover the molecular evolutionary mechanisms driving structural adaptations of the vestibular system in these animals, we conducted an extensive study. Our research focused on 42 mammalian species, spanning 42 genera, 20 orders, and 38 families, and encompassed 12 semi-aquatic species alongside 30 terrestrial counterparts. Furthermore, we curated a list of 116 genes known to influence the development or functionality of the vestibular system through an exhaustive review of pertinent literature. This gene set can also serve as a reference for other similar studies, such as investigating the genetic basis of sense of balance evolution in mammals with other modes of locomotion (e.g., flight). It may even provide insights for medical fields, such as the treatment of vertigo.

Evolution of genes involved in semicircular canals development

The functional interplay between the overall size and morphological diversity of the semicircular canal and locomotor behaviors stands as a well-established phenomenon [1, 6]. Previous investigations have delineated significant morphological disparities in semicircular canals between semi-aquatic and terrestrial mammals [8, 10-12]. Through our analysis, we have identified 27 genes potentially pivotal for the adaptive evolution of the sense of balance in semi-aquatic mammals. Among these genes, 48.1% (13/27) — ADGRG6, CASP3, CHD7, COG4, GATA2, GSDME, HDAC1, ILDR1, MYCN, OTX1, SLC26A2, SOX10, and TSKU - are implicated in the development of semicircular canals. Of particular significance is the SLC26A2 gene, notable for its involvement in positive selection, divergent evolution, rapidly evolved, semi-aquatic correlation, and convergent evolution. Similarly, the SOX10 gene, categorized as both a divergent evolution gene and a gene with semi-aquatic correlation. These two genes emerge as potential candidates



Fig. 5 The interaction network between proteins of 21 genes. Among the 27 candidate genes, there are significant interactions between the proteins of 21 genes (*p* < 1.0e-16). Each node in the network represents the protein of a gene. The filling in the nodes represents the 3D structure of the protein. The differently colored connecting lines between nodes represent different types of interaction evidence: "_____" for from database, "____" for experimental validation, "____" for text mining, "____" for gene neighborhood, "____" for gene fusions, "____" for gene co-occurrence, "____" for co-expression, and "____" for protein homology

for regulating the diverse morphology of semicircular canals in semi-aquatic mammals during their adaptation to aquatic locomotion. Of the three semicircular canals, special attention is focused on the horizontal semicircular canal, which is closely associated with motor agility [8]. The *MYCN* and *OTX1* genes investigated in this study play a crucial role in the normal development of the horizontal semicircular canals. Given their correlation with semi-aquatic traits and rapid evolutionary rates, these genes may govern the development of the horizontal semicircular canal, thus enhancing the precise perception of semi-aquatic locomotor behaviors in semi-aquatic mammals.

The genes identified in association with semicircular canal development offer molecular insights into the morphological diversity of these canals in response to alterations in locomotor behaviors. Nevertheless, the complex morphogenesis of inner ear semicircular canals necessitates meticulous regulation, involving processes like cell proliferation and epithelial-mesenchymal transition [48]. The quest for identifying the precise regulatory mechanisms requires further investigation within the fields of genetics and physiology.

Evolution of genes involved in otolith development

In the utricle and the saccule, the two otolith organs, the presence of denser otoliths heightens sensitivity to gravitational forces and linear acceleration. This increased sensitivity is due to the inertia of otoliths, which resist movement and exert a shearing force on stationary cilia. This force triggers nerve impulses within the vestibulocochlear nerve [50]. Despite extensive comparative morphological analyses of the semicircular canals, there has been limited attention given to comparative anatomical investigations of mammalian otolithic organs. Prior research is constrained, with only one study highlighting the significantly thicker otolithic membranes in cetaceans compared to terrestrial and semi-aquatic mammals. However, this research lacks quantitative data and overlooks proportional effects [51].

In the present study, examination of 27 genes revealed that 25.9% (7/27) of them (HDAC1, NOXO1, OC90, OTOP1, SIX1, SLC26A2, and SLC26A4) are involved in otolith formation. Remarkably, among these genes, SLC26A2 was consistently present in all five datasets, while the OTOP1 gene appeared in four datasets simultaneously, and the OC90 gene was also found concurrently in two datasets. The identification of adaptive evolutionary genes associated with otoliths suggests that otoliths may also have undergone selective pressures during the evolutionary process of mammalian locomotion. This implies that otoliths might have experienced modifications in their sensitivity to better suit a semi-aquatic lifestyle. Future research endeavors could explore the specific influence of these genes on otolith perception across different locomotor modalities.

Evolution of genes associated with vestibular disorders

A comprehensive investigation into the intricate molecular and cellular mechanisms governing vestibular morphogenesis is indispensable not only for elucidating the development of this complex organ but also for comprehending the etiology of vestibular abnormalities [52]. Within our study, we have discerned that 18.5% (5/27) of the genes (CDH23, GJB2, GJB6, KCNA10, and USH1C) constitute the genetic underpinning for vestibular disorders in humans. Notably, two of these genes (GJB6 and USH1C) were concurrently identified in three datasets. The CDH23 and USH1C genes are recognized as the causal agents for the USH1C and USH1D subtypes of Usher syndrome type I (USH1), respectively. It is important to note that individuals with USH1 often exhibit vestibular dysfunction, with delays in motor development serving as a clinical hallmark of congenital vestibular impairments [38]. In contradistinction to terrestrial animals, which navigate both terrestrial and aquatic terrains, semi-aquatic mammals confront the unique challenge of traversing both land and underwater domains in three dimensions. This necessitates the maintenance of stability during activities such as foraging or evading predators. Consequently, the vestibular system of semi-aquatic mammals is subjected to distinct demands to uphold physical balance [53]. We posit that the distinctive evolutionary trajectories of the USH1C and CDH23 genes lay the groundwork for the adaptation of intricate locomotor patterns in semi-aquatic mammals. Understanding the mechanisms of these genes in semi-aquatic species could inform potential treatments for patients with USH1.

In instances of DFNB1 deafness stemming from mutations in GJB2 and/or GJB6, the majority of patients manifest vestibular dysfunction. For many, alleviating vertigo requires reclining, while others endure vertigo severe enough to disrupt their daily activities [38]. Additionally, irregularities in vestibular otolith fragments can trigger the pathological sensation of vertigo [20]. Vertigo, characterized by the perception of motion in oneself or the environment, is a prevalent disorder of the human vestibular system. Approximately 7.4% of adults experience lifelong bouts of vertigo, significantly hampering their work performance and daily functionality, thus posing a considerable burden on healthcare systems [54]. Furthermore, beyond terrestrial habitats, semi-aquatic mammals, such as otter species like sea otters, partake in rapid rotational movements underwater [55]. Understanding the mechanisms employed by these creatures to manage vertigo holds promising implications for the advancement of treatment strategies for human vertigo.

Conclusion

Multiple mammalian clades have independently evolved to thrive in semi-aquatic environments. We investigated the evolutionary mechanisms of sense of balance related genes in these species, crucial for maintaining locomotive balance between land and water. Our dataset, comprising 116 genes linked to the vestibular system, was analyzed across 12 semi-aquatic and 30 terrestrial mammalian species. Our findings propose that *SLC26A2* and SOX10 are key candidates for morphological changes in the semicircular canals, crucial for the evolution of the vestibular system in response to shifts in motor patterns. These genes, in conjunction with *MYCN* and *OTX1*, likely orchestrate semicircular canal formation to adapt to semi-aquatic habitats. Additionally, genes involved in otolith development, such as *SLC26A2*, *OC90*, and *OTOP1*, might modulate otolith sensory acuity and other functions across diverse movement modalities. Genes like *GJB2*, *GJB6*, and *USH1C*, linked to vestibular disorders, could constitute the molecular underpinnings enabling semi-aquatic mammals to mitigate vertigo amidst intricate movement patterns.

Methods

Species coverage and construction of gene set

Within the Mammalia class, there are approximately 140 species that exhibit a semi-aquatic lifestyle [50, 53]. In our study, we selected 12 semi-aquatic mammal species and 30 terrestrial mammal species for our research based on the publication status of semi-aquatic mammalian genomes, the availability of species genomes in our laboratory, the quality of genome assembly, and the phy-logenetic distribution of the species. We have reviewed extensive literature and compiled nearly all genes known prior to the commencement of this study that are associated with the development of the vestibular system (semicircular canals, otoliths, and vestibular structures). These genes will be used as a gene set for subsequent analyses.

Sequence acquisition and construction of the phylogenetic tree

Among the 42 selected species (refer to Additional file 1: Table S7 for genomic details), genome sequencing was conducted in our laboratory for the *Aonyx cinereus*, *Lutra lutra*, *Neovison vison*, *Martes flavigula*, and *Vulpes ferrilata*. However, the genome sequences of the *Aonyx cinereus*, *Lutra lutra*, *Neovison vison* are not yet publicly available. Genome data for the remaining species were acquired from the NCBI database (https://www.ncbi. nlm.nih.gov/).

For annotated genomes, protein-coding sequences of the target genes were directly retrieved from the NCBI database. In cases where multiple protein-coding sequences were available for a gene, we selected the longest sequence after alignment. For unannotated genomes and target sequences not found in annotated genomes, we utilized the target gene sequences of *Homo sapiens*, *Mus musculus*, *Equus caballus*, *Manis javanica*, and *Enhydra lutris* as query sequences. Utilizing TBtools software [56] and GeneWise (https://www.ebi.ac.uk/ Tools/psa/genewise/), we obtained the protein coding sequences of these target genes. Afterwards, we preserved only the longest transcript for each gene. In the BLAST process, we set a stringent E-value threshold of 1e-10. In cases where multiple homologous sequences were still present, we included them in the dataset for clustering analysis to ensure that the correct sequences were identified. Additionally, we conducted a bidirectional BLAST analysis using the RBH (Reciprocal Best Hit) method [57] to verify whether the genes we obtained are orthologous. Based on the identified occurrences of the target genes across the 42 species, we carried out multiple bidirectional BLAST analyses on various combinations of species datasets.

To facilitate subsequent analyses of molecular evolution, correlation, and convergent evolution, we concatenated the orthologous genes identified in all 42 species in the same order. We then performed sequence alignment using the PRANK program [58]. To improve the quality of the alignment, we subsequently trimmed the sequence alignment results using the Gblocks program [59], eliminating regions with poor alignment quality. Finally, we used IQ-TREE (-m GTR+F+I+G4 -bb 1000 -alrt 1000) and MrBayes (ngen=1,000,000, samplefreq=1000, nchains=4) to construct the maximum likelihood (ML) and Bayesian inference (BI) trees, respectively.

Selection pressure analysis

To assess the impact of natural selection on genes associated with the sense of balance, we employed the codeml program within the PAML software package [60]. Initially, we utilized the branch-site model (Ma: model=2, NSsites=2, fix_omega=0; null model Ma0: model=2, NSsites = 2, fix_omega = 1, omega = 1) to identify potential positive selection among genes shared within semi-aquatic mammal subgroups. In this analysis, semiaquatic species were designated as foreground branches, contrasting with terrestrial species as background branches. Each gene underwent thorough examination, with loci possessing a posterior probability (BEB) exceeding 0.8 considered indicative of positive selection [61]. This approach led to the identification of genes harboring at least one such locus as positively selected genes shared by the semi-aquatic group.

Concurrently, site model (M2a: model=0, NSsites=2; null model M1a: model=0, NSsites=1. M8: model=0, NSsites=8; null model M7: model=0, NSsites=7) analysis was independently conducted for each gene across all terrestrial mammals. This method aimed to isolate positively selected loci, thereby minimizing the influence of genes exhibiting positive selection across all species and facilitating the identification of genes specifically favored in semi-aquatic species.

Subsequently, to identify genes under divergent evolution and those that have rapidly evolved in semi-aquatic species following their adaptation to aquatic habitats, all species were categorized into semi-aquatic and terrestrial groups. We applied Clade model C (CmC) [62] (CmC: model=3, NSsites=2; null model M2a_rel: model=0, NSsites=22) to calculate the evolutionary rate for each gene within these groups. Genes displaying significantly different evolutionary rates between the two control groups were designated as divergent evolution genes. Additionally, we utilized the Branch model (M2: model=2, NSsites=0; null model M0: model=0, NSsites = 0) to analyze all genes within the two groups. Genes exhibiting a significantly higher evolutionary rate compared to the terrestrial group were classified as rapidly evolved genes shared among the semi-aquatic group. Finally, the free ratio model (M1: model = 1, NSsites = 0; null model M0: model=0, NSsites=0) in the branch model was employed to ascertain the respective evolutionary rates of genes across all species for subsequent correlation analysis.

Investigating the correlation between gene evolutionary rates and habitats

To explore whether the evolutionary rates of different genes are linked to specific locomotor patterns in various habitats, we conducted correlation analyses using Phylogenetic Independent Contrast (PIC) methods [63]. PIC analyses are instrumental in mitigating phylogenetic biases when assessing diverse traits across species. Initially, we logarithmically adjusted the evolutionary rate of each gene along the terminal branches, as determined by the free ratio model. Subsequently, we categorized terrestrial and semi-aquatic species as 0 and 1, respectively, for correlation analyses between these two subgroups. The phylogenetic tree with branch lengths utilized in this study was obtained from the TimeTree database. All analyses were executed in R using the ape package [64], with specific commands detailed in accordance with the procedures outlined on website https://www.r-phylo.org/ wiki/HowTo/Phylogenetic_Independent_Contrasts.

Convergent evolutionary analysis

Semi-aquatic mammals confront the dual challenge of adapting to two distinct locomotor modes: terrestrial movement and aquatic swimming, stemming from their secondary aquatic adaptation. Our investigation aims to discern whether species with independently evolving semi-aquatic adaptations share similar molecular traits in maintaining their sense of balance. We conducted an analysis of convergent evolution at the protein level employing the PCOC (Profile Change with One Change) method, which has exhibited superior efficacy in identifying convergence compared to other approaches. In this approach, convergence at a site is delineated as a shift from the biochemical properties of an ancestral branch amino acid to an amino acid with distinct biochemical properties shared by multiple convergent clades. Essentially, our objective was to discern convergence in amino acid properties as opposed to mere identical amino acid substitutions [65, 66]. PCOC operates as a phylogeny-based examination utilizing branch lengths derived from the TimeTree database. Semi-aquatic species are delineated as evolutionarily discrete entities based on their phylogenetic ties and are allocated to distinct clades, each representing independent secondary aquatic adaptations. These branches encompass: clade 1 (Condylura cristata), clade 2 (Enhydra lutris, Lutra lutra, Aonyx cinereus, and Neovison vison), clade 3 (Zalophus californianus, Odobenus rosmarus divergens, and Phoca vitulina), clade 4 (Ursus maritimus), clade 5 (Hippopotamus amphibius), clade 6 (Castor canadensis), and clade 7 (Hydrochoerus hydrochaeris). We set a posteriori probability threshold of 0.95 for the identification of convergent loci, categorizing genes manifesting at least one convergent locus as convergent evolutionary genes.

Protein interaction analysis

To investigate the protein interactions among positively selected genes, rapidly evolved genes, genes with semiaquatic correlation, and convergent evolution genes obtained from the previous analysis, we utilized the STRING v12.0 database (https://string-db.org/). This database is specifically designed to gather, assess, and consolidate all publicly available information on protein interactions, supplementing it with computational predictions [67]. By using the STRING database, we can investigate whether there are significant interactions among the candidate genes obtained from the analysis results, to substantiate the validity of our findings.

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s12862-024-02345-9.

Additional file 1: Table S1. List of related genes for sense of balance and references. Table S2. The version number of the sequence downloaded from the NCBI database. Table S3. List of positively selected genes in terrestrial group mammals (site model: comparison between M8 model and M7 model). Table S4. List of positively selected genes in terrestrial group mammals (site model: comparison between M2 model and M1 model). Table S5. List of divergent evolution genes in semi-aquatic group mammals (Clade model C). Table S6. List of rapidly evolving genes in semi-aquatic group mammals (Branch model). Table S7. Information of 42 mammal genomes used in this study.

Additional file 2. The raw data of all sequences obtained in this study.

Additional file 3. The results of bidirectional BLAST with the RBH method on all identified orthologous gene sequences.

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Authors' contributions

Y.D.: Conceptualization, Data curation, Investigation, Writing—original draft. Q.W.: Conceptualization, Funding acquisition, Supervision. G.S. and X.G.: Investigation, Writing—review & editing. T.L. and L.W.: Validation, Resources. S.Z. and X.W.: Software, Methodology. Y.S. and L.S.: Investigation, Software, Data curation. H.Z.: Conceptualization, Funding acquisition, Project administration. All authors read and approved the final manuscript.

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Data availability

The datasets generated and analyzed during the current study are available in Additional file 2 in Supplementary Information.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

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References

- 1. Ekdale EG. Form and function of the mammalian inner ear. J Anat. 2016;228(2):324–37.
- Graf WM. Evolution of the vestibular system. In: Binder MD, Hirokawa N, Windhorst U, editors. Encyclopedia of neuroscience. Berlin Heidelberg: Springer; 2009. p. 1440-1448.
- Latimer AE, Sherratt E, Bonnet T, Scheyer TM. Semicircular canal shape diversity among modern lepidosaurs: life habit, size, allometry. BMC Ecology and Evolution. 2023;23(1):10.
- Mackowetzky K, Yoon KH, Mackowetzky EJ, Waskiewicz AJ. Development and evolution of the vestibular apparatuses of the inner ear. J Anat. 2021;239(4):801–28.
- Davies KTJ, Bates PJJ, Maryanto I, Cotton JA, Rossiter SJ. The evolution of bat vestibular systems in the face of potential antagonistic selection pressures for flight and echolocation. PLoS ONE. 2013;8(4):e61998.
- Bronzati M, Benson RBJ, Evers SW, Ezcurra MD, Cabreira SF, Choiniere J, Dollman KN, Paulina-Carabajal A, Radermacher VJ, Roberto-da-Silva L, et al. Deep evolutionary diversification of semicircular canals in archosaurs. Curr Biol. 2021;31(12):2520-2529.e2526.
- Pfaff C, Martin T, Ruf I. Bony labyrinth morphometry indicates locomotor adaptations in the squirrel-related clade (Rodentia, Mammalia). Proceedings of the Royal Society B-Biological Sciences. 1809;2015(282):20150744.
- Grohe C, Tseng ZJ, Lebrun R, Boistel R, Flynn JJ. Bony labyrinth shape variation in extant Carnivora: a case study of Musteloidea. J Anat. 2016;228(3):366–83.
- Spoor F, Bajpai S, Hussain ST, Kumar K, Thewissen JGM. Vestibular evidence for the evolution of aquatic behaviour in early cetaceans. Nature. 2002;417(6885):163–6.

- 10. Mara Loza C, Latimer AE, Sanchez-Villagra MR, Carlini AA. Sensory anatomy of the most aquatic of carnivorans: the Antarctic Ross seal, and convergences with other mammals. Biol Let. 2017;13(10):20170489.
- 11. Schwab JA, Young MT, Neenan JM, Walsh SA, Witmer LM, Herrera Y, Allain R, Brochu CA, Choiniere JN, Clark JM, et al. Inner ear sensory system changes as extinct crocodylomorphs transitioned from land to water. Proc Natl Acad Sci USA. 2020;117(19):10422–8.
- 12. Ekdale EG. Comparative anatomy of the bony labyrinth (inner ear) of placental mammals. PLoS ONE. 2013;8(6):e66624.
- Georgi JA, Sipla JS. 15 Comparative and Functional Anatomy of Balance in Aquatic Reptiles and Birds. In: Thewissen J, Nummela S, editors. Sensory Evolution on the Threshold: Adaptations in Secondarily Aquatic Vertebrates. Berkeley: University of California Press; 2008. p. 233-256.
- 14. Alsina B, Whitfield TT. Sculpting the labyrinth: morphogenesis of the developing inner ear. Semin Cell Dev Biol. 2017;65:47–59.
- Requena T, Espinosa-Sanchez JM, Lopez-Escamez JA. Genetics of dizziness: cerebellar and vestibular disorders. Curr Opin Neurol. 2014;27(1):98–104.
- 16. Chatterjee S, Kraus P, Lufkin T. A symphony of inner ear developmental control genes. BMC Genet. 2010;11:68.
- Hwang CH, Keller J, Renner C, Ohta S, Wu DK: Genetic interactions support an inhibitory relationship between bone morphogenetic protein 2 and netrin 1 during semicircular canal formation. Development 2019, 146(4):dev174748.
- Mazan S, Jaillard D, Baratte B, Janvier P. OTX1 gene-controlled morphogenesis of the horizontal semicircular canal and the origin of the gnathostome characteristics. Evol Dev. 2000;2(4):186–93.
- Pauley S, Wright TJ, Pirvola U, Ornitz D, Beisel K, Fritzsch B. Expression and function of FGF10 in mammalian inner ear development. Developmental Dynamics: an Official Publication of the American Association of Anatomists. 2003;227(2):203–15.
- 20. Dror AA, Taiber S, Sela E, Handzel O, Avraham KB. A mouse model for benign paroxysmal positional vertigo with genetic predisposition for displaced otoconia. Genes Brain Behav. 2020;19(5):e12635.
- Kozel PJ, Friedman RA, Erway LC, Yamoah EN, Liu LH, Riddle T, Duffy JJ, Doetschman T, Miller ML, Cardell EL, et al. Balance and hearing deficits in mice with a null mutation in the gene encoding plasma membrane Ca2+-ATPase isoform 2. J Biol Chem. 1998;273(30):18693–6.
- Duncan JS, Stoller ML, Francl AF, Tissir F, Devenport D, Deans MR. CELSR1 coordinates the planar polarity of vestibular hair cells during inner ear development. Dev Biol. 2017;423(2):126–37.
- Du Y, Ren L, Jiang Q, Liu X, Ji F, Zhang Y. Yuan S-I, Wu Z-M, Guo W-W, Yang S-M: Degeneration of saccular hair cells caused by MITF gene mutation. Neural Dev. 2019;14(1):1.
- 24. He Y, Tang D, Li W, Chai R, Li H. Histone deacetylase 1 is required for the development of the zebrafish inner ear. Sci Rep. 2016;6:16535.
- Zou D, Erickson C, Kim E-H, Jin D, Fritzsch B, Xu P-X. EYA1 gene dosage critically affects the development of sensory epithelia in the mammalian inner ear. Hum Mol Genet. 2008;17(21):3340–56.
- Liu F, Xia W, Hu J, Wang Y, Yang F, Sun S, Zhang J, Jiang N, Wang H, Tian W, et al. Solute carrier family 26 member a2 (SLC26A2) regulates otic development and hair cell survival in zebrafish. PLoS ONE. 2015;10(9):e0136832.
- Armstrong PA, Wood SJ, Shimizu N, Kuster K, Perachio A, Makishima T. Preserved otolith organ function in caspase-3-deficient mice with impaired horizontal semicircular canal function. Exp Brain Res. 2015;233(6):1825–35.
- Clement A, Blanco-Sanchez B, Peirce JL, Westerfield M. COG4 is required for protrusion and extension of the epithelium in the developing semicircular canals. Mech Dev. 2019;155:1–7.
- Elmaleh-Berges M, Baumann C, Noel-Petroff N, Sekkal A, Couloigner V, Devriendt K, Wilson M, Marlin S, Sebag G, Pingault V. Spectrum of temporal bone abnormalities in patients with Waardenburg Syndrome and SOX10 mutations. Am J Neuroradiol. 2013;34(6):1257–63.
- Miwa T, Ito N, Ohta K. Tsukushi is essential for the formation of the posterior semicircular canal that detects gait performance. Journal of Cell Communication and Signaling. 2021;15(4):581–94.
- Kiss PJ, Knisz J, Zhang YZ, Baltrusaitis J, Sigmund CD, Thalmann R, Smith RJH, Verpy E, Banfi B. Inactivation of NADPH oxidase organizer 1 - results in severe imbalance. Curr Biol. 2006;16(2):208–13.

- Petko JA, Millimaki BB, Canfield VA, Riley BB, Levenson R. OTOC1: a novel otoconin-90 ortholog required for otolith mineralization in zebrafish. Dev Neurobiol. 2008;68(2):209–22.
- Hurle B, Ignatova E, Massironi SM, Mashimo T, Rios X, Thalmann I, Thalmann R, Ornitz DM. Non-syndromic vestibular disorder with otoconial agenesis in tilted/mergulhador mice caused by mutations in OTOPETRIN 1. Hum Mol Genet. 2003;12(7):777–89.
- Macova I, Pysanenko K, Chumak T, Dvorakova M, Bohuslavova R, Syka J, Fritzsch B, Pavlinkova G. NEUROD1 is essential for the primary tonotopic organization and related auditory information processing in the midbrain. J Neurosci. 2019;39(6):984–1004.
- Riccomagno MM, Takada S, Epstein DJ. Wht-dependent regulation of inner ear morphogenesis is balanced by the opposing and supporting roles of SHH. Genes Dev. 2005;19(13):1612–23.
- 36. Zine A, Aubert A, Qiu J, Therianos S, Guillemot F, Kageyama R, de Ribaupierre F. HES1 and HES5 activities are required for the normal development of the hair cells in the mammalian inner ear. The Journal of Neuroscience: the Official Journal of the Society for Neuroscience. 2001;21(13):4712–20.
- Yan D, Liu XZ. Genetics and pathological mechanisms of Usher syndrome. J Hum Genet. 2010;55(6):327–35.
- Dodson KM, Blanton SH, Welch KO, Norris VW, Nuzzo RL, Wegelin JA, Marin RS, Nance WE, Pandya A, Arnos KS. Vestibular dysfunction in DFNB1 deafness. Am J Med Genet A. 2011;155A(5):993–1000.
- Lee SI, Conrad T, Jones SM, Lagziel A, Starost MF, Belyantseva IA, Friedman TB, Morell RJ. A null mutation of mouse KCNA10 causes significant vestibular and mild hearing dysfunction. Hear Res. 2013;300:1–9.
- Lentz J, Pan F, Ng SS, Deininger P, Keats B. USH1C216A knock-in mouse survives Katrina. Mutation Research-Fundamental and Molecular Mechanisms of Mutagenesis. 2007;616(1–2):139–44.
- Swanson WJ, Vacquier VD. The rapid evolution of reproductive proteins. Nat Rev Genet. 2002;3(2):137–44.
- 42. Chai S, Tian R, Bi J, Xu S, Yang G, Ren W. Rapid evolution and molecular convergence in cryptorchidism-related genes associated with inherently undescended testes in mammals. BMC Ecology and Evolution. 2021;21(1):22.
- 43. Geng F-S, Abbas L, Baxendale S, Holdsworth CJ, Swanson AG, Slanchev K, Hammerschmidt M, Topczewski J, Whitfield TT. Semicircular canal morphogenesis in the zebrafish inner ear requires the function of gpr126 (lauscher), an adhesion class G protein-coupled receptor gene. Development. 2013;140(21):4362–74.
- 44. Sang Q, Zhang J, Feng R, Wang X, Li Q, Zhao X, Xing Q, Chen W, Du J, Sun S, et al. Ildr1b is essential for semicircular canal development, migration of the posterior lateral line primordium and hearing ability in zebrafish: implications for a role in the recessive hearing impairment DFNB42. Hum Mol Genet. 2014;23(23):6201–11.
- Haugas M, Lillevali K, Hakanen J, Salminen M. GATA2 is required for the development of inner ear semicircular ducts and the surrounding perilymphatic space. Dev Dyn. 2010;239(9):2452–69.
- Dominguez-Frutos E, Lopez-Hernandez I, Vendrell V, Neves J, Gallozzi M, Gutsche K, Quintana L, Sharpe J, Knoepfler PS, Eisenman RN, et al. N-MYC controls proliferation, morphogenesis, and patterning of the inner ear. J Neurosci. 2011;31(19):7178–89.
- 47. Shah AM, Krohn P, Baxi AB, Tavares ALP, Sullivan CH, Chillakuru YR, Majumdar HD, Neilson KM, Moody SA. SIX1 proteins with human branchio-oto-renal mutations differentially affect cranial gene expression and otic development. Disease Models & Mechanisms. 2020;13:3.
- Hurd EA, Micucci JA, Reamer EN, Martin DM. Delayed fusion and altered gene expression contribute to semicircular canal defects in CHD7 deficient mice. Mech Dev. 2012;129(9–12):308–23.
- Busch-Nentwich E, Sollner C, Roehl H, Nicolson T. The deafness gene DFNA5 is crucial for UGDH expression and HA production in the developing ear in zebrafish. Development. 2004;131(4):943–51.
- Davis RW. Marine mammals: adaptations for an aquatic life. 1st ed. Cham (Switzerland) and New York: Springer; 2019.
- Spoor F, Thewissen JGM. 16 Comparative and Functional Anatomy of Balance in Aquatic Mammals. In: Thewissen J, Nummela S, editors. Sensory Evolution on the Threshold: Adaptations in Secondarily Aquatic Vertebrates. Berkeley: University of California Press; 2008. p. 257-284.

- Rakowiecki S, Epstein DJ. Divergent roles for Wnt/β-catenin signaling in epithelial maintenance and breakdown during semicircular canal formation. Development. 2013;140(8):1730–9.
- 53. Hood GA: Semi-aquatic mammals: ecology and biology: Johns Hopkins University Press; 2020.
- 54. Eppsteiner RW, Smith RJH. Genetic disorders of the vestibular system. Curr Opin Otolaryngol Head Neck Surg. 2011;19(5):397–402.
- Kenyon KW. The sea otter in the eastern Pacific Ocean. 1st ed. US Government Printing Office Washington: US Bureau of Sport Fisheries and Wildlife; 1969.
- Chen C, Chen H, Zhang Y, Thomas HR, Frank MH, He Y, Xia R. TBtools: an integrative toolkit developed for interactive analyses of big biological data. Mol Plant. 2020;13(8):1194–202.
- Moreno-Hagelsieb G, Latimer K. Choosing BLAST options for better detection of orthologs as reciprocal best hits. Bioinformatics. 2008;24(3):319–24.
- Loytynoja A, Goldman N. Phylogeny-aware gap placement prevents errors in sequence alignment and evolutionary analysis. Science. 2008;320(5883):1632–5.
- Castresana J. Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. Mol Biol Evol. 2000;17(4):540–52.
- 60. Alvarez-Carretero S, Kapli P, Yang Z. Beginner's Guide on the Use of PAML to Detect Positive Selection. Mol Biol Evol. 2023;40:4.
- Yang Z, Wong WSW, Nielsen R. Bayes empirical bayes inference of amino acid sites under positive selection. Mol Biol Evol. 2005;22(4):1107–18.
- Weadick CJ, Chang BSW. An improved likelihood ratio test for detecting site-specific functional divergence among clades of protein-coding genes. Mol Biol Evol. 2012;29(5):1297–300.
- 63. Felsenstein J. Phylogenies and the comparative method. Am Nat. 1985;125(1):1–15.
- 64. Paradis E, Claude J, Strimmer K. APE: analyses of phylogenetics and evolution in R language. Bioinformatics. 2004;20(2):289–90.
- Rey C, Gueguen L, Semon M, Boussau B. Accurate detection of convergent amino-acid evolution with PCOC. Mol Biol Evol. 2018;35(9):2296–306.
- Potter JHT, Drinkwater R, Davies KTJ, Nesi N, Lim MCW, Yohe LR, Chi H, Zhang X, Levantis I, Lim BK, et al. Nectar-feeding bats and birds show parallel molecular adaptations in sugar metabolism enzymes. Curr Biol. 2021;31(20):4667-4674.e4666.
- Szklarczyk D, Gable AL, Lyon D, Junge A, Wyder S, Huerta-Cepas J, Simonovic M, Doncheva NT, Morris JH, Bork P, et al. STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. Nucleic Acids Res. 2019;47(D1):D607–13.

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